



Compact Dry™ TC

Ready-to-Use Medium for
Total Viable Count



Background

It is important to detect and measure the total viable count in food products and the food environment to monitor the degree of cleanliness as well as sanitary safety. The pour plate method has been widely used to determine microbial counts. The pour plate method is time-consuming and complicated, requiring operations such as the preparation of hot agar maintained at 45–50°C, and uniform mixing and dilution. To save operator time and make it possible for anyone to perform the microbial count test without difficulty, Compact Dry was developed based on a new concept and technology applicable to the food industry. Compact Dry allows for easy addition of a sample to the device.

Certification by AOAC

Compact Dry TC has been compared to AOAC Official MethodSM 966.23 and certified by the AOAC Research Institute Performance Tested MethodsSM Program (Certificate No. 010404) for enumeration of total viable counts in raw meat (raw ground beef, raw ground pork, raw pork, raw lamb, and raw veal). A matrix extension comparing Compact Dry TC to ISO 4833:2003 for cooked chicken, fresh pre-washed bagged shredded iceberg lettuce, frozen cod filets, instant nonfat dry milk, and pasteurized 2% milk was approved in 2015. A matrix extension to raw chicken breast with enumeration at 24 hours and 48 hours and a modification for raw ground beef for enumeration at 24 hours compared to FSIS MLG 3.02 were approved in 2020.

Warnings and Precautions

1. General precautions

- Read and follow precisely the warnings and directions for use described in the package insert and/or label.
- Do not use the product after its expiration date. Quality of the product is not warranted after its shelf life expires.
- Do not use product that contains any foreign materials, is discolored or dehydrated, or has a damaged container.
- Use plates as soon as possible after opening. Return any unused plates to the aluminum bag and seal with tape to avoid light and moisture. Compact Dry TC is sensitive to light, which affects the color development of colonies.
- Cap tightly after inoculation to avoid dehydration of gelled medium.

2. Safety precautions

- If medium or reagent comes into contact with eyes or mouth, immediately wash with water and consult a physician.
- Procedures with microorganisms involve certain risks of laboratory-acquired infections. Procedures should be carried out under the supervision of trained laboratory personnel with biohazard protection measures.
- Treat any laboratory equipment or medium that comes into contact with the specimen as infectious and sterilize appropriately.

3. Precautions for disposal of waste

- Sterilize any medium, reagent or materials by autoclaving or boiling after use, and then dispose of it as industrial waste according to local laws and regulations for disposal of such material.

4. User responsibilities

- It is the user's responsibility in selecting any test method to evaluate a sufficient number of samples with particular foods and microbial challenges to satisfy the user that the chosen test method meets the user's criteria.
- It is the user's responsibility to determine that any test methods and results meet its customers or suppliers' requirements. The user must train its personnel in proper testing techniques.
- It is the user's responsibility to validate the performance of this method for use with any non-certified matrix.

5. Limitation of warranties

- Compact Dry plates are manufactured at ISO 9001:2015 facility. If any Compact Dry plate is proven to be defective by fault of the manufacturer or its authorized distributors, they may replace or, at their discretion, refund the purchase price of any plate. These are the exclusive remedies.

Storage and Shelf Life

Storage: Keep at room temperature (1–30°C)

Shelf life: Twenty-four (24) months after manufacturing. Expiration date is printed on outer box label and aluminum bag label.

Package

Compact Dry TC 100 plates	Code 54051
Compact Dry TC 1400 plates	Code 54051-CS

Further Information

Customer Support

Shimadzu Diagnostics Corporation
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**Kit components, operating
instructions and interpretation**



Test Kit Components

1. Compact Dry TC Plates

Additional Reagents and Supplies Required, Not Provided

1. Butterfield's phosphate-buffered diluent (BPBD) — Prepare according to AOAC 966.23
2. Maximum recovery diluent (MRD) — Prepare according to ISO 4833:2003
3. Filtered Stomacher bags

Apparatus

1. Blender or Stomacher or equivalent for homogenizing sample
2. Pipets: 1 ml
3. Incubator: 35 ± 1°C (raw meat products) or 30 ± 1°C (all other matrices)

Operating Procedure

Preparation of specimen

1. **Prepare appropriate diluent:** Butterfield's phosphate-buffered diluent (BPBD) for raw meat products or maximum recovery diluent (MRD) for other claimed matrices. Autoclave for sterilization.
2. **Viable count in solid food products:** For raw meat, weigh 50 g of sample and add 450 ml BPBD to the sample. Homogenize by blender for 2 min ± 15s. For cooked chicken, fresh lettuce, or frozen fish, weigh 10 g of sample and add 90 ml MRD. Homogenize by Stomacher for 1 min ± 10s. For milk powder, weigh 10 g of sample and add to 90 ml MRD pre-warmed to 45 ± 1°C. Slowly swirl and shake until sample is dissolved.
3. **Viable count in liquid food products:** For pasteurized milk, use without dilution, dilute 1 ml in 9 ml MRD, or dilute further if viable count is >300 cfu/plate. Vortex to mix.
4. **Viable count in swab test sample** (not included in AOAC PTM certification):
Wiping solution, which is obtained from cotton swab, is used without dilution or diluted in MRD. It is recommended to use Swab Test ST-25PBS (Code 06698) available as an optional kit.

Directions for Compact Dry TC

1. Open an aluminum bag and take out a set of four plates.
2. Detach the necessary number of plate(s) from a set of four by bending up and down while pressing the lid. Use a set of four connected plates when serial dilution measuring is intended.
3. Remove cap from plate, pipette 1 ml of sample (to be diluted further if necessary) in the middle of the dry plate and replace cap. Specimen diffuses automatically and evenly over the entire plate (total medium of 20 cm²) to transform it into a gel within seconds.
4. Write the appropriate sample information in the memorandum section. Invert the capped plate and place in incubator at 35 ± 1°C for raw meat or 30 ± 1°C for all other matrices. Incubate 48 ± 3 h.
5. From the backside of the plate, count the number of colonies (colored and colorless) in the medium. White paper placed under the plate can make colony counting easier. For large numbers of colonies, use the grids carved on the backside consisting of 1 cm x 1 cm, or 0.5 cm x 0.5 cm, at the four corners.
6. Enumeration range of Compact Dry TC is 1–300 cfu/plate. Specimens should be diluted in buffer to obtain a concentration level less than 300 cfu/plate.

Precautions for Use

1. Do not use Compact Dry TC for human and animal diagnosis.
2. To avoid microbial contamination, do not touch the surface of the dry plate medium during inoculation.
3. During incubation, keep cap tight to avoid any possible dehydration.
4. Use of filtered Stomacher bags is recommended to eliminate risks of carryover of tiny pieces of foodstuffs onto the surface of the medium.
5. If more than 10⁴ CFU/ml were inoculated onto a plate, no distinguishable colored colonies will form and the entire plate will become colored.
6. If the nature of the sample affects the reaction of the medium, inoculate the sample only after the factor has been eliminated by means such as dilution, pH adjustment or other. This may include samples with high viscosity, that are colored, that react with the redox indicator, or that have too high or too low pH.

Interpretation

1. The medium consists of non-selective medium and the redox indicator 2,3,5-Triphenyl Tetrazolium Chloride (TTC). Colonies grown on Compact Dry TC are almost all red colored.
2. The full plate size is 20 cm². The backside contains carved grids of 1 cm x 1 cm and 0.5 cm x 0.5 cm to make colony counting easier. If large numbers of colonies are present on the medium, the total viable count can be obtained by averaging the number of colonies per large grid (1 cm x 1 cm), counted from several grids, and multiplying by 20. Alternatively, the total viable count can be obtained by averaging the number of colonies per small grid (0.5 cm x 0.5 cm), counted from several grids, and multiplying by 80.
3. Since some microorganisms may not reduce TTC to develop red/pink color, colonies may develop on Compact Dry TC that are not necessarily red. All colonies should be counted.